

## UKVDL Customer Testing Information

### Procedures for BOVINE TRICHOMONIASIS SAMPLE COLLECTION AND SUBMISSION

The diagnosis of Trichomoniasis venereal infection in cattle is contingent upon proper sample collection submission in the appropriate transport medium. **Note: If testing is required for interstate movement, check with the state of destination for the specific test requirements.**

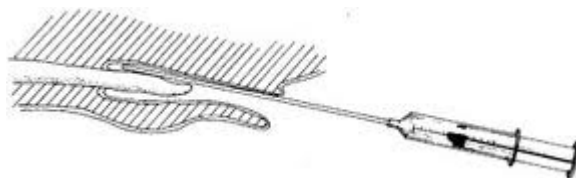
#### Materials required for sample collection and submission:

1. Sampling device: one for each animal to be sampled. You will need a 28 inch (71.72 cm) long catheter with an outside dimension of 0.215 inches (0.546 cm), assembled with a 19 inch (48.26 cm) tube protector.
2. Transport media: *Trichomonas foetus*–samples for PCR should be submitted in InPouch TF pouches, trich tube (with transport media) or direct smegma in PBS or sterile saline (5 ml). For culture, samples must be submitted in InPouch™ TF pouches.
3. Other items: Disposable gloves (one pair for each animal sampled), Sterile Saline, sterile 12 cc syringes, single use paper towels.

#### Collection procedures: Male animals

The organism inhabits the preputial cavity and epithelial crypts of the glans penis. Sexual rest for 1-2 weeks is required before sampling bulls.

1. Restrain animal adequately.
2. Use a separate pair of gloves and a separate collection device for each animal.
3. Clean debris from the preputial orifice and clip preputial hairs to about one-half inch length. If necessary, rinse out the preputial cavity with sterile saline to clean out mud and manure. This will help prevent contamination from non-pathogenic intestinal trichomonads.
4. Insert the sampling device (pipette inside tube or plastic sheath) into the preputial opening to about the distal third of the preputial cavity.
5. Advance the collecting pipette through and beyond the protecting tube to the preputial fornix.



6. Collect the sample by rapidly scraping the pipette back and forth in short strokes on the mucosa of the distal penis and fornix area while applying suction with a rubber bulb or syringe and massaging the glans penis through the sheath to move smegma into the pipette. Fifteen to thirty (15-30) strokes of the pipette are required to obtain an adequate sample.
7. Retract the pipette back into the protecting tube and remove the entire device from the preputial cavity.

8. Collect at least one inch (2.54 cm) of smegma in the end of the collecting pipette. Do not get blood.

### **Collection procedures: Female animals**

The organisms are found in the cervical mucus. If cervical mucus or vaginal discharge cannot be obtained, the anterior vagina may be sampled.

1. Restrain animal adequately.
2. Use a separate pair of gloves and a separate collection device for each animal.
3. Clean debris from the vulva.
4. Immobilize the cervix per rectum and insert the sampling device into the anterior third of the vagina.
5. Pass the collection pipette through the protecting tube and advance it to the cervical os.
6. Apply suction with a rubber bulb or syringe to aspirate cervical mucus into the pipette. Some persistence may be required to aspirate the thick mucus from this area. If postcoital pyometra due to *T. foetus* is suspected, the medium should be inoculated with a specimen of the uterine exudate.
7. Retract the pipette into the retracting tube and remove the entire collection device from the vagina.

### **Inoculation of the InPouch™ Transport Media**

Remove the pouch from the bag and, if necessary, manually express the liquid from one chamber to the other, resulting in approximately 1 ml in the upper chamber. Be sure the liquid in the upper chamber is below the closure tape to prevent fluid from leaking when the pouch is opened. Tear open the pouch at the notch located just above the closure tape. Open the pouch by pulling the closure tapes middle tabs apart. Insert the specimen pipette tip into the liquid of the pouch's upper chamber and expel 0.5-1.0 cc of the sample into the pouch. If the collected material adheres to the wall of the pipette, rinse the pipette by flushing a small amount of the liquid medium back and forth into the pouch. Minimize the production of bubbles. Squeeze the top to close, roll the top to close, roll the top edge down and continue rolling twice. Fold the wire tabs over to prevent the InPouch™ from reopening.

### **Shipping Requirements**

- Complete the Accession Form and specify test for Trichomoniasis PCR or Culture. Send samples to the UKVDL as soon as possible following collection. The laboratory should receive the samples within 24-48 hours after collection. Label all samples with the date of collection and official animal identification.
- Package the samples so they are protected from extreme heat and cold. Samples should be maintained at room temperature (65-80 °F, 18-27 °C) while in transit. Clients should schedule shipments to avoid weekend and holiday delivery of samples to the laboratory.